

Hepatotoxic impact of desloratadine/dihydroartemisinin/piperaquine on healthy and parasitized mice

Georgewill UO¹, Adikwu E², Ebong NO^{1*}

To Cite:

Georgewill UO, Adikwu E, Ebong NO. Hepatotoxic impact of desloratadine/dihydroartemisinin/piperaquine on healthy and parasitized mice. *Drug Discovery*. 2022; 16(37), 36-44

Author Affiliation:

¹Department of Pharmacology, Faculty of Basic Clinical Sciences, University of Port Harcourt, Rivers State, Nigeria

²Department of Pharmacology and Toxicology, Faculty of Pharmacy, Niger Delta University, Bayelsa State, Nigeria.

***Corresponding author:**

Email: nwakaepong@gmail.com
+2348132321014

Peer-Review History

Received: 02 February 2022

Reviewed & Revised: 06/February/2022 to 05/April/2022

Accepted: 06 April 2022

Published: 09 April 2022

Peer-review

External peer-review was done through double-blind method.



© The Author(s) 2022. Open Access. This article is licensed under a [Creative Commons Attribution License 4.0 \(CC BY 4.0\)](http://creativecommons.org/licenses/by/4.0/), which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

ABSTRACT

Background: Desloratadine/dihydroartemisinin/piperaquine (DL/D/P) can be used for malaria treatment, but its safety assessment is imperative. **Objective:** To evaluate its hepatotoxic effect on healthy and *Plasmodium berghei*-infected mice. **Method:** Fifty-four adult Swiss albino mice (25-30g) were used. The mice, n=6/group were inoculated with *Plasmodium berghei* (1×10^7) and treated with DL (5mg/kg), D/P (1.71/13.7mg/kg) and DL/D/P daily for 4 days, respectively. The healthy mice were treated with DL (5mg/kg), D/P (1.71/13.7mg/kg) and DL/D/P daily for 28 days, respectively. After drug treatment, the mice were weighed and anesthetized. Blood samples were collected and assessed for liver function indices. Liver samples were excised, weighed and evaluated for oxidative stress markers and histology. **Results:** DL, D/P and DL/D/P had no significant ($p>0.05$) effects on liver function parameters in parasitized mice when compared to control. DL, D/P and DL/D/P significantly decreased body weight and significantly increased liver weight in healthy mice at $p<0.05$, $p<0.05$, and $p<0.01$, respectively when compared to control. Serum aminotransferases, gamma-glutamyl transferase, lactate dehydrogenase, alkaline phosphatase and bilirubin levels increased significantly while total protein and albumin levels decreased significantly in healthy mice treated with DL ($p<0.05$), D/P($p<0.01$) and DL/D/P ($p<0.001$) when compared to control. Significantly decreased liver catalase, glutathione peroxidase superoxide dismutase, and glutathione with significantly increased malondialdehyde levels were observed in healthy mice treated with DL ($p<0.05$), D/P ($p<0.01$) and DL/D/P ($p<0.001$) when compared to control. DL/D/P produced hepatocyte necrosis in healthy mice. **Conclusion:** Malaria treatment with DL/D/P may be safe on the liver, but prolonged use may cause liver dysfunction.

Keywords: Dihydroartemisinin/piperaquine, Desloratadine, *Plasmodium*, Hepatotoxicity, Mice

1. INTRODUCTION

The liver is a vital organ responsible for an array of functions that aids metabolism, immunity, bile secretion, digestion, detoxification, and vitamin

storage (Karla *et al.*, 2020). The liver can thus be regarded as an organ involved in sustaining and regulating homeostasis in the body. Almost all biochemical pathways to growth, energy provision, nutrient supply, and reproduction requires the liver (Sharma *et al.*, 1991). The Liver's anatomy and physiology enables it function in the disposition of drugs administered orally, serving as a portal to tissues and a major site of drug metabolism (Baillie and Rettie, 2011). Drug metabolism by the liver sometimes produces active and toxic metabolites which may cause hepatotoxicity (Remmer, 1970).

Hepatotoxicity associated with some antimalarial drugs has raised some notable concern (Omotuyi *et al.*, 2008; Owumi *et al.*, 2015; Yin *et al.*, 2014). Studies in humans have shown elevation of liver enzymes of clinical significance (Ribeiro, and Olliaro, 1998). Severe hepatotoxicity with visible systemic signs has been reported with the use of chloroquine (Farver and Lavin, 1999; Lee, 2003; Liu, 2015). A clear correlation between antimalarial drug administration and elevated liver enzymes has been observed in some artemisinin-based combination therapies (ACTs), (Moore, 2018). Halofantrine was reported to cause elevations of liver enzymes and pathologic changes in guinea pigs, such as severe hepatic degeneration (Obi *et al.*, 2004).

Dihydroartemisinin-piperaquine (D/P) is part of the five ACTs currently recommended by the World Health Organization (WHO) for the treatment of uncomplicated malaria infection (Zani *et al.*, 2014). Piperaquine which has a half-life of several weeks has activity against chloroquine-resistant *Plasmodium vivax* and *Plasmodium falciparum* (Hung *et al.*, 2004; Tarning *et al.*, 2008). Dihydroartemisinin is the active metabolite of artemether and artesunate. D/P is an effective and frequently used ACT (Gutman *et al.*, 2017). It may be safe (Myint *et al.*, 2007), but reports suggest it may cause hepatic dysfunction (Okafor, Ufele and Nwankwo, 2019; Batty *et al.*, 2008; Mesembe *et al.*, 2009). Desloratadine (DL) is a non-sedating second generation H1-antihistamine that was established for the treatment of allergic rhinitis in 2001 (Villa, 2001), but studies showed it has potential antimalarial activity (Aneesa, 2011) and showed synergistic activity with chloroquine (Aneesa, 2011). In recent studies, DL increased the antimalarial activity of D/P in *Plasmodium berghei*-infected mice, (Georgewill *et al.*, 2021), but with a paucity of scientific information on the safety of their combination especially on the liver. Hence the present study assessed the hepatotoxic effect of combined DL and D/P on healthy and *Plasmodium berghei*-infected mice which is imperative.

2. MATERIALS AND METHODS

2.1. Animals, malaria parasite and drugs

A total of fifty-four adult Swiss albino mice (n=6/group) of both sexes (25-30g) obtained and kept at the animal house of the Department of Pharmacology, Faculty of Basic Clinical Sciences, University of Port Harcourt, Rivers State, Nigeria were used. The mice were kept in well ventilated and clean cages, maintained under standard environmental conditions and fed with standard laboratory animal food pellets with water *ad libitum*. DL (Merck & Co) and D/P (Bliss GVS Pharma Ltd, India) were used. The following doses from previous studies on the antiplasmodial activity of DL/D/P were used: D/P (1.71/13.7 mg/kg) and DL (5 mg/kg) (Georgewill *et al.*, 2021). Donor mice infected with Chloroquine-sensitive strain of *Plasmodium berghei* (*P. berghei*) (NK65) provided by the Nigerian Institute of Medical Research (NIMR), Yaba, Lagos, Nigeria were used. The donor mice were sacrificed and blood samples were collected by cardiac puncture. The blood samples were diluted to 2ml with 0.9% saline containing 1×10^7 parasitized erythrocytes and were used to intraperitoneally (ip) infect the experimental mice used.

2.2. Parasite inoculation, treatment and animal sacrifice

Thirty mice randomized into n=6/ group were used. The mice were grouped II-V and inoculated (i.p) with *P. berghei* containing 1×10^7 parasitized erythrocytes. After 3 days, treatment commenced orally as follows: Group 1(Normal control) and group II (parasitized control) received 0.2 ml of normal saline, groups III-V received (D/P) (1.71/13.7 mg/kg), DL (5 mg/kg) and DL/D/P daily for 4 days, respectively. For the sub-acute study, twenty-four healthy mice of n=6/group of four groups were used. Group I (Control) daily received 0.2ml of normal saline orally for 28 days. Groups II-IV orally received (D/P) (1.71/13.7 mg/kg), DL (5 mg/kg) and DL/D/P daily for 28 days, respectively. After drug administration, the mice were fasted overnight, weighed and anesthetized with diethyl ether and blood samples were obtained by cardiac puncture. Blood samples were centrifuged at a speed of 1200 rpm for 20 minutes and sera separated and evaluated for biochemical markers. Mice were dissected and the liver harvested and rinsed in saline. The rinsed liver were homogenized in 0.1 M Tris-HCl solution buffered (pH 7.4) and centrifuged (2000 rmp for 20 minutes). The supernatants were decanted and evaluated for oxidative stress markers.

2.3. Serum biochemical marker assessments

2.3.1 Assessments of liver biochemical markers

Serum alanine aminotransferase (ALT), total bilirubin (TB), alkaline phosphatase (ALP), aspartate aminotransferase (AST), albumin, and lactate dehydrogenase (LDH) were measured using laboratory test apparatus according to the manufacturer's specifications.

2.3.2 Evaluation of oxidative stress markers

Liver glutathione (GSH) was assayed as described by Sedlak and Lindsay (1968). Superoxide dismutase (SOD) was estimated as reported by Sun and Zigman (1978). Catalase (CAT) was measured using the procedure explained by Aebi (1984). Glutathione peroxidase (GPx) was determined as explained by Rotruck *et al.*, (1973). Malondialdehyde (MDA) was measured according to the method described by Buege and Aust (1978).

2.4. Histology of the liver

Liver tissues were harvested and preserved in 10% buffered formalin for 24hr. The Liver tissues were dehydrated in graded alcohol concentrations. The tissues were processed, embedded in paraffin wax and sectioned (3 µm each). The sectioned liver tissues were stained with Eosin and Hematoxylin and examined with a light microscope.

2.5. Statistical analysis

Data were expressed as mean ± SEM. Data were subjected to one-way Analysis of Variance (ANOVA) and complemented with Tukey's multiple range test using Graph Pad Prism 5 Software (San Diego, CA USA). Statement on statistical significance was based on p< 0.05; p<0.01 and p<0.001.

3. RESULTS

Effects of desloratadine/dihydroartemisinin/piperaquine on body and liver weights of healthy and *Plasmodium berghei*-infected Mice

Body and liver weights were not altered (p>0.05) in *P. berghei*-infected mice treated with DL, D/P and DL/D/P for 4 days, respectively when compared to control (Table 1). Following 28 days of treatment, body weight significantly decreased while liver weight significantly increased in healthy mice treated with DL (p<0.05), D/P (p<0.05) and DL/D/P (p<0.01), respectively when compared to control (Table 1).

Table 1: Effect of desloratadine/dihydroartemisinin/piperaquine on body and liver weights of healthy and *Plasmodium berghei*-infected mice

Treatment	Final body weight (g)		Absolute liver weight (g)		Relative liver weight (%)	
	Healthy mice	Parasitized mice	Healthy mice	Parasitized mice	Healthy mice	Parasitized mice
Control	30.40±2.27	25.00±2.28	1.83±0.06	1.85±0.03	6.02±0.03	7.40±0.57
DL	25.43±2.21*	24.80±3.12	2.33±0.05*	1.83±0.07	9.16±0.65*	7.37±0.34
D/P	24.20±2.69*	24.40±2.78	2.30±0.04*	1.80±0.09	9.50±0.72*	7.38±0.02
DL/ D/ P	21.70±2.85 ^π	23.21±3.47	2.69±0.06 ^π	1.77±0.04	12.40±1.07 ^π	7.63±0.06

Data as mean ± SEM, SEM: Standard error of mean. n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, * p<0.05, ^π p<0.01

Significant difference when compared to control (Healthy mice)

Effects of desloratadine/dihydroartemisinin/piperaquine on serum liver biochemical markers of healthy and *Plasmodium berghei*-infected mice

Serum AST, ALT, ALP, LDH, TB, total protein and albumin levels remained unchanged (p>0.05) in *P. berghei*-infected mice treated with DL, D/P and DL/D/P for 4 days, respectively when compared to control (Table 2). However, significant increases in serum AST, ALT, ALP, LDH, and TB levels with significant decreases in serum total protein and albumin levels were detected in healthy mice treated with DL (p<0.05), D/P (p<0.01) and DL/D/P (p<0.001), respectively when compared to control (Table 3). In the healthy mice, treatment with DL, D/P and DL/D/P produced no significant changes (p>0.05) on serum TG, CH, HDL-C, and LDL-C levels when compared to control (Table 4).

Table 2: Effect of desloratadine/dihydroartemisinin/piperaquine on serum liver biochemical markers of *Plasmodium berghei*-infected mice

Treatment	ALT (U/L)	AST (U/L)	ALP (U/L)	LDH (U/L)	TB (g/dL)	T. Protein (g/dL)	Albumin (g/dL)
Control	30.20±2.21	38.60±4.48	27.80±4.07	33.05±3.12	3.14±0.37	5.66±0.20	4.46±0.06
PC	31.50±2.34	39.20±4.65	28.30±4.12	36.17±3.65	3.22±0.47	5.43±0.42	4.40±0.08
DL	29.60±3.15	38.70±4.35	29.20±3.07	34.50±2.12	3.37±0.53	5.38±0.32	4.37±0.12
D/P	29.80±2.57	40.40±4.57	29.60±4.28	37.50±4.32	3.35±0.31	5.32±0.67	4.34±0.05
DL/ D/ P	31.20±2.38	43.80±4.18	30.20±3.65	38.90±4.15	3.50±0.23	5.21±0.20	4.30±0.04

Data as mean± SEM, n=6, SEM: Standard error of mean, PC: Parasitized untreated control, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, AST: Aspartate aminotransferase, ALT: Alanine aminotransferase, LDH: Lactate dehydrogenase, TB: Total bilirubin, ALP: Alkaline phosphatase, T. Protein: Total protein

Table 3: Effect of desloratadine/dihydroartemisinin/piperaquine on serum liver biomarkers of healthy mice

Treatment	ALT (U/L)	AST (U/L)	ALP (U/L)	LDH (U/L)	TB (g/dL)	T. Protein (g/dL)	Albumin (g/dL)
Control	31.73±3.03	43.80±6.27	25.6±3.21	33.10±3.17	3.12±0.32	8.30±0.12	5.34±0.45
DL	37.60±4.12*	65.70±7.04*	39.12±4.34*	60.32±4.60*	5.68±0.61	6.31±0.06*	3.71±0.09*
D/P	60.21±5.29**	89.21±6.57**	58.20±6.23**	79.20±5.31**	6.10±0.71**	6.00±0.21**	3.20±0.09**
DL/ D/ P	98.10±6.23 ^π	163.9±8.23 ^π	101.20±8.03 ^π	113.7±7.25 ^π	8.01±0.42 ^π	4.50±0.84 ^π	2.01±0.64 ^π

Data as mean± SEM, n=6, SEM: Standard error of mean, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, AST: Aspartate aminotransferase, ALT: Alanine aminotransferase, LDH: Lactate dehydrogenase, TB: Total bilirubin, ALP: Alkaline phosphatase, T. Protein: Total protein, * p<0.05, **p<0.01 and ^πp<0.001 Significant difference when compared to control.

Table 4: Effect of desloratadine/dihydroartemisinin/piperaquine on serum lipid parameters of healthy mice

Treatment	T g/dL	CHOL g/dL	HDL-C g/dL	LDL-C g/dL
Control	51.60±5.32	87.50±8.51	31.70±3.21	28.60±3.56
DL	52.90±6.37	89.70±7.04	30.90±3.07	29.10±3.01
D/P	55.40±7.20	93.20±9.05	32.50±3.71	30.40±4.37
DL/ D/ P	57.20±7.90	97.40±9.11	34.70±3.11	31.30±4.16

Data as mean± SEM, n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, T: Triglyceride, CHOL: Total cholesterol, HDL-C: High density lipoprotein cholesterol, LDL-C: Low density lipoprotein cholesterol. SEM: Standard error of mean.

Table 5: Effect of desloratadine/dihydroartemisinin/piperaquine on liver oxidative stress markers of *Plasmodium berghei*-infected mice

Treatment	MDA nmole/mg protein	GSH μmole/mg protein	CAT U/mg protein	SOD U/mg protein	GPx U/mg protein
Control	0.24±0.05	16.30±1.44	25.33±3.07	20.23±3.04	15.29±1.56
PC	0.25±0.07	16.27±0.98	25.30±2.55	20.11±2.88	15.22±1.99
DL	0.23±0.09	16.26±1.23	25.27±3.77	20.09±3.22	15.19±1.77
D/P	0.25±0.06	16.23±1.45	24.90±4.32	19.88±1.31	15.00±1.63
DL/ D/ P	0.26±0.07	16.20±1.11	24.80±2.46	19.67±1.52	14.97±1.21

Data as mean ± SEM, SEM: Standard error of mean, PC: Parasitized untreated control, DL: Desloratadine, D/P:

Dihydroartemisinin/piperaquine, MDA: Malondialdehyde, GSH: Glutathione, CAT: Catalase, SOD: Superoxide dismutase GPx: Glutathione peroxidase.

Table 6: Effect of desloratadine/dihydroartemisinin/piperaquine on liver oxidative stress markers of healthy mice

Treatment	MDA nmole/mg protein	GSH μmole/mg protein	CAT U/mg protein	SOD U/mg protein	GPx U/mg protein
Control	0.23±0.01	16.71±0.64	27.10±1.07	17.11±2.04	17.81±0.80
DL	0.37±0.03*	12.40±0.71*	22.61±3.56*	13.47±2.43*	13.90±1.02*
D/P	0.49±0.05**	8.72±0.55**	18.21±2.61**	11.23±1.53**	10.50±0.31**
DL/ D/ P	0.61±0.04 ^π	5.27± 0.37 ^π	11.70±1.35 ^π	7.07±2.81 ^π	7.60±0.41 ^π

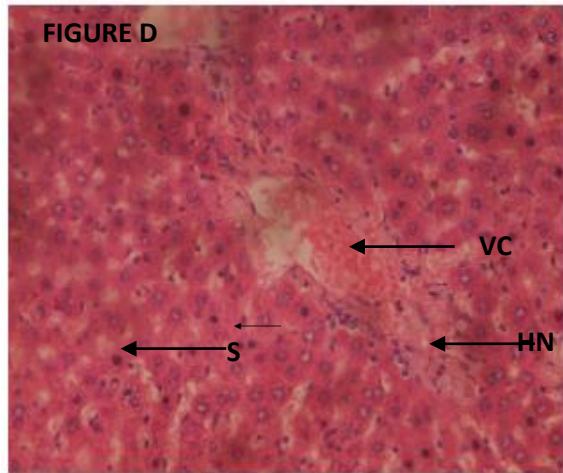
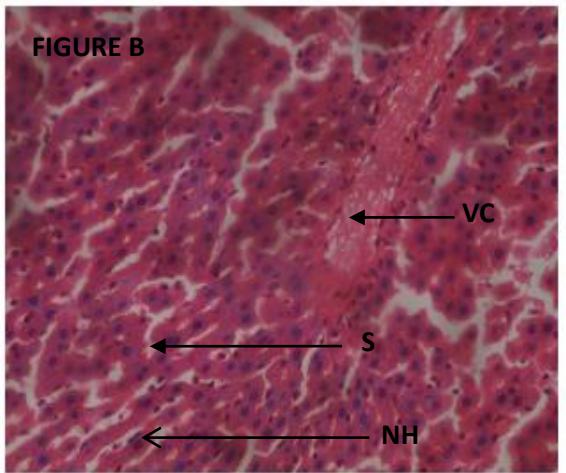
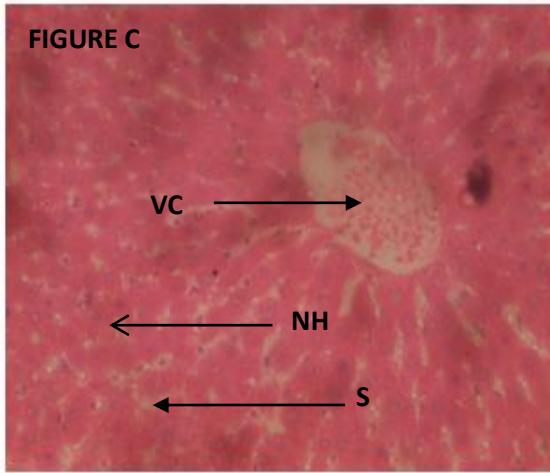
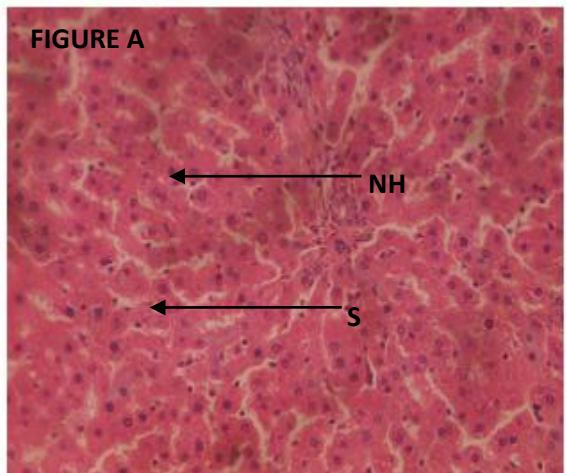
Data as mean ± SEM, SEM: Standard error of mean, n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, D/P/DL: MDA:

Malondialdehyde, GSH: Glutathione, CAT: Catalase, SOD: Superoxide dismutase GPx: Glutathione peroxidase, * p<0.05, **p<0.01, ^π

p<0.001 Significant difference when compared to control.

Effects of desloratadine/dihydroartemisinin/piperaquine on liver oxidative stress markers and histology of healthy mice

The activities of liver antioxidants (SOD, GSH, CAT and GPx) were normal in *P. berghei*-infected mice treated for 4 days (Table 5), but were significantly decreased in healthy mice treated with DL ($P < 0.05$), D/P ($P < 0.01$) and DL/D/P ($P < 0.001$) for 28 days when compared to control (Table 6). MDA levels were normal in *P. berghei*-infected mice treated for 4 days (Table 6), but were significantly increased in healthy mice following 28 days treatment with DL ($P < 0.05$), D/P ($P < 0.01$) and DL/D/P ($P < 0.001$) when compared to control (Table 6). The photomicrographs of the liver of the control mice showed normal hepatocytes and central vein (Figure A). The liver of DL -treated healthy mice showed central vein congestion (Figure B). The liver of D/P-treated healthy mice showed central vein congestion (Figure C). The liver of DL/D/P-treated healthy mice showed central vein congestion and hepatocyte necrosis (Figure D).



The liver of the control mice (Figure A), the liver of DL-treated healthy mice (Figure B), the liver of D/P-treated healthy mice (Figure C) and the liver of DL/D/P-treated healthy mice (Figure D) X 400. NH: Normal hepatocytes, VC: Central vein congestion HN: Hepatocyte necrosis, S: Sinusoids

4. DISCUSSION

Studies on toxicity are important steps in the development of new drugs (Parasuraman, 2011). Considering the potential antimalarial benefit of DL/D/P, comprehensive knowledge on its toxicity profile is lacking especially on the liver. The present study evaluated the hepatotoxicity of DL/D/P in healthy and *P. berghei*-infected mice. The assessment of body and organ weights in toxicity studies is an essential step in the evaluation of chemical substances (Sellers *et al.*, 2007). An abnormal change in an organ weight caused by an administered xenobiotic is an indicator of its toxicity (Teo *et al.*, 2002; Wang *et al.*, 2007). In this present study, DL/D/P had no effects on the liver and body weights of *P. berghei*-infected mice treated for 4 days. But decreased body weight and increased liver weight occurred in healthy mice treated with DL/D/P for 28 days. In healthy mice, it implies that DL/D/P may have decreased appetite and induced inflammation in the liver of treated mice. Liver biochemical markers evaluated in this study are important and effective parameters used for the diagnoses of liver diseases (Adikwu *et al.*, 2020). In this study, treatment with DL/D/P had no negative effect on serum liver biochemical markers of *P. berghei*-infected mice treated for 4 days. However, treatment of healthy mice for 28 days with DL/D/P altered serum liver biochemical markers marked by increased serum AST, ALT, ALP, TB, LDH levels and decreased total protein and albumin levels. The observation is a sign of hepatic damage (Adikwu *et al.*, 2020), which may be caused by the distortion of hepatocyte membrane leading to the leakage of hepatocyte cytosolic contents (Bhattacharyya, 2003). There were no observable changes in serum T, CHL, LDL-C and HDL-C levels of DL/D/P-treated healthy rats for 28 days.

Oxidative stress is a consequence of the overproduction of ROS by metabolic reactions associated with oxygen, which alters oxidant/antioxidant balance in favour of the oxidant (Betteridge, 2000; Birben *et al.*, 2012). ROS are unpaired electrons and highly reactive molecules, when in excess they react with various biological biomolecules in cells (nucleic acids, lipids, and proteins) causing functional and structural damage (Birben *et al.*, 2012). In this study, liver oxidative stress makers of *P. berghei*-infected mice treated with DL/D/P were not altered. In healthy mice, DL/D/P altered liver oxidative stress markers marked by decreased antioxidants and elevated MDA level. Free radical-induced lipid peroxidation, plays an important function in pathological processes. Free radical-induced lipid peroxidation can be measured by conjugated dienes, MDA, and 4-hydroxynonenal, but MDA has been frequently used (Grotto *et al.*, 2009). DL/D/P had no conspicuous effect on MDA level of *P. berghei*-infected mice treated for 4 days. In contrast, DL/D/P visibly increased MDA levels in healthy mice treated for 28 days. This indicates that DL/D/P caused hepatic lipid peroxidation in healthy mice. This may be attributed to the induction of excess ROS production in the liver by DL/D/P leading to oxidative stress. The liver of DL/D/P treated healthy mice for 28 days showed hepatic necrosis and central vein congestion. This finding could be due to liver bimolecular damage to DNA, proteins, lipids and other cellular components through oxidative stress caused by ROS production (Grotto *et al.*, 2009).

5. CONCLUSION

Results from this study showed that treatment of malaria using DL/D/P may not cause hepatotoxicity except with long term use.

Acknowledgement

Drug administration and animal handling performed by the Staff of Pharmacology Animal House, Faculty of Basic Clinical Sciences, University of Port Harcourt, Nigeria are acknowledged.

Funding:

This study did not receive any external funding.

Ethical approval

The Animal ethical guidelines are followed in the study for experimentation.

Conflict of Interest:

The authors declare that there are no conflicts of interests.

Data and materials availability:

All data associated with this study are present in the paper.

REFERENCES

- Adikwu E, Ebinyo NC, Benalayefa O. Protective effect of lycopene against tamoxifen-induced hepatotoxicity in albino rats. *Biomedical and Biotechnology Research Journal*. 2020, 4, 69-75
- Aebi H. Catalase in vitro. *Methods Enzymol*. 1984, 105, 121-126
- Aneesa S. Evaluation of antihistamines for in vitro antimalarial activity against Plasmodium falciparum. 2001, <https://api.semanticscholar.org/CorpusID:82705763>. Accessed 29 October 2020
- Baillie TA, Rettie AE. Role of biotransformation in drug-induced toxicity: influence of intra- and inter-species differences in drug metabolism. *Drug Metab Pharmacokinet*. 2011, 26(1), 15-29
- Batty KT, Moore BR, Stirling V, Ilett KF, Page-Sharp M, Shilkin KB, Mueller I, Karunajeewa HA, Davis TME. Toxicology and pharmacokinetics of piperaquine in mice. *Toxicology*. 2008, 249, 55-61
- Betteridge DJ. What is oxidative stress? *Metabolism*. 2000, 49(2 Suppl 1), 3-8
- Bhattacharya U, Roy S, Kar PK, Sarangi B, Lahiri SC. *Indian J Med Res*. 1988, 88, 558–563
- Bhattacharyya D, Mukherjee R, Pandit S, Das N, Sur TK. Prevention of carbon tetrachloride induced hepatotoxicity in rats by Himoliv®, a polyherbal formulation. *Ind. J. Pharmacol*. 2003, 35, 183-185
- Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. *World Allergy Organ J*. 2012, 5(1), 9-19
- Björnsson ES. Hepatotoxicity by Drugs: The Most Common Implicated Agents. *Int J Mol Sci*. 2016, 17(2), 224

11. Buege JA, Aust SD. Microsomal lipid peroxidation. *Meth Enzymol.* 1978, 52, 302-10
12. Enonwu CO, Afolabi BM, Salako LO, Idigbe EO, Bashirelahi N. *J Neural Transm.* 2000, 107, 1273–1287
13. Farver DK, Lavin MN. "Quinine-induced hepatotoxicity." *The Annals of Pharmacotherapy.* 1999, 33(1), 32–34
14. Gbotosho GO, Happi CT, Ganiyu A, Ogundahunsi OA, Sowunmi A, Oduola AM. Potential contribution of prescription practices to the emergence and spread of chloroquine resistance in south-west Nigeria: caution in the use of artemisinin combination therapy. *Malar J.* 2009, 8, 313
15. Georgewill UO, Ebong NO, Adikwu E. Antiplasmodial activity of desloratadine-dihydroartemisinin-piperaquine on Plasmodium berghei infected mice. *Journal of Applied Biology and Biotechnology.* 2021, 9(2), 169-173
16. Grotto D, Maria LM, Valentini J, Paniz C, Schmitt GS, Garcia SC, Pomblum VJ, Rocha JBT, Farina M. Importance of the lipid peroxidation biomarkers and methodological aspects FOR malondialdehyde quantification. *Quim. Nova.* 2009, 32(1), 169-174
17. Gutman J, Kovacs S, Dorsey G, Stergachis A, Ter Kuile FO. Safety, tolerability, and efficacy of repeated doses of dihydroartemisinin-piperaquine for prevention and treatment of malaria: a systematic review and meta-analysis. *Lancet Infect Dis.* 2017, 17(2), 184-193
18. Gutteridge JMC. Lipid peroxidation and antioxidants as biomarkers of tissue damage. *Clin. Chem.* 1995, 41, 1819 – 1828
19. Hung TY, Davis TM, Ilett KF, Karunajeewa H, Hewitt S, Denis MB, Lim C, Socheat D. Population pharmacokinetics of piperaquine in adults and children with uncomplicated falciparum or vivax malaria. *Br J Clin Pharmacol.* 2004, 57(3), 253-62
20. Kalra A, Yetiskul E, Wehrle CJ, Tuma F. Physiology, Liver. In: *StatPearls.* Treasure Island (FL): StatPearls Publishing; May 9, 2021.
21. Kenilworth NJ. Desloratadine (AERIUS®): Prescribing information. Schering Plough Corporation, 2005
22. Kharchoufa L, Bouhrim M, Bencheikh N, et al. Acute and Subacute Toxicity Studies of the Aqueous Extract from *Haloxylon scoparium* Pomel (*Hammada scoparia* (Pomel)) by Oral Administration in Rodents. *Biomed Res Int.* 2020, 2020, 4020647
23. Lee W. "Drug-induced hepatotoxicity." *The New England Journal of Medicine.* 2003, 349, 474–485
24. Liu AC. "Hepatotoxic reaction to chloroquine phosphate in a patient with previously unrecognized porphyria cutanea tarda." *Western Journal of Medicine.* 1995, 162(6), 548–551
25. Maegraith B, Fletcher A. The pathogenesis of mammalian malaria. *Adv Parasitol.* 1972, 10, 49-75
26. Mesembe OE, Ekam VS, Mfem CC, Igiri AO, Ekanem TB. Modulatory Effect of Vitamin C and E on Dihydroartemisinin (Cotecxin) Induced Hepatotoxicity in Wistar Albino Rats. *Mary Slessor Journal of Medicine.* 2009, 9(1)
27. Moore BR. Liver injury in uncomplicated malaria: an overlooked phenomenon. *EBioMedicine.* 2018, 37, 15-16
28. Myint HY, Ashley EA, Day NP, Nosten F, White NJ. Efficacy and safety of dihydroartemisinin-piperaquine. *Trans R Soc Trop Med Hyg.* 2007, 101(9), 858-866
29. Obi E, Orisakwe OE, Asomugha LA, Udemezue OO, Orish VN. The hepatotoxic effect of halofantrine in guinea pigs. The hepatotoxic effect of halofantrine in guinea pigs. *Indian J Pharmacol.* 2004, 36(5), 303-305
30. Okafor UE, Ufele AN, Nwankwo OD. Effects of artemisinin-based combination therapy on histopathology of the liver, kidney and spleen of mice infected with Plasmodium berghei. *Animal Research International.* 2019, 16(3), 3519 – 3528
31. Olayinka ET, Ore A. Alterations in Antioxidant Status and Biochemical Indices Following Administration of Dihydroartemisinin-Piperaquine Phosphate (P-ALAXIN®). *IOSR Journal of Pharmacy and Biological Sciences.* 2013, 5(4), 43–53
32. Omotuyi IO, Nwangwu SC, Okugbo OT, Okoye OT, Ojeh GC, Wogu DM. Hepatotoxic and hemolytic effects of acute exposure of rats to artesunate overdose. *Afr J Biochem Res.* 2008, 2, 107–10
33. Owumi SE, Gbadegesin MA, Odunola OA, Adegoke AM, Uwaifo AO. Toxicity associated with repeated administration of artemether-lumefantrine in rats. *Environ Toxicol.* 2015, 30, 301–7
34. Pandit A, Sachdeva T, Bafna P. Drug-Induced Hepatotoxicity: A Review. *Journal of Applied Pharmaceutical Science.* 2012, 02 (05), 233-243
35. Parasuraman S. Toxicological screening. *Journal of Pharmacology & Pharmacotherapeutics.* 2011, 2(2), 74–79
36. Remmer H. The role of the liver in drug metabolism. *Am J Med.* 1970, 49, 617-629
37. Ribeiro IR, Olliari P. Safety of artemisinin and its derivatives: a review of published and unpublished clinical trials. *Med Trop (Mars).* 1998, 58, 50–3
38. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Sci.* 1973, 179, 588-590
39. Sedlak J, Lindsay RH. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal Biochem.* 1968, 25, 192-205
40. Sharma A, Chakraborti KK, Handa SS. Anti-hepatotoxic activity of some Indian herbal formulations as compared to silymarin. *Fitoterapia.* 1991, 62, 229-235

41. Singhal A, Morris VB, Labhasetwar V, Ghorpade A. Nanoparticle-mediated catalase delivery protects human neurons from oxidative stress. *Cell Death Dis.* 2013, 4(11), e903
42. Srichaikul T, Archararit N, Siriasawakul T, Viriyapanich T. *Trans R Soc Trop Med Hyg.* 1976, 70, 36–38
43. Sun M, Zigma S. An Improved spectrophotometer assay of superoxide dismutase based on epinephrine antioxidation. *Anal Biochem.* 1978, 90, 81-9
44. Sureshkumar D, Begum S, Johannah NM, Maliakel B, Krishnakumar IM. Toxicological evaluation of a saponin-rich standardized extract of fenugreek seeds (FenuSMART): acute, sub-chronic and genotoxicity studies. *Toxicology Reports.* 2018, 5, 1060–1068
45. Tarning J, Ashley EA, Lindegarth N, Stepniewska K, Phaiphun L, Day NP, McGready R, Ashton M, Nosten F, White NJ. Population pharmacokinetics of piperaquine after two different treatment regimens with dihydroartemisinin-piperaquine in patients with Plasmodium falciparum malaria in Thailand. *Antimicrob Agents Chemother.* 2008, 52(3), 1052-61
46. Teo S, Stirling D, Thomas S, Hoberman A, Kiorpis A, Khetani V. A 90-day oral gavage toxicity study of D-methylphenidate and D,L-methylphenidate in Sprague-Dawley rats. *Toxicology.* 2002, 179(3), 183–196
47. Villa E, Rogkakou A, Garelli V, Canonica W. Review of Desloratadine Data Using the ARIA Guidelines. *World Allergy Organ J.* 2012, 5, S6–S13
48. Wang TC, Su YP, Hsu TY, Yang CC, Lin CC. "28-Day oral toxicity study of the aqueous extract from spider brake (*Pteris multifida* Poiret) in rats." *Food and Chemical Toxicology.* 2007, 45(9), 1757–1763
49. Yin JY, Wang HM, Wang QJ, Dong YS, Han G, Guan YB, Zhao KY, Qu WS, Yuan Y, Gao XX, Jing SF, Ding RG. Subchronic toxicological study of two artemisinin derivatives in dogs. *PLoS One.* 2014, 9, e94034
50. Zani B, Gathu M, Donegan S, Olliaro PL, Sinclair D. Dihydroartemisinin-piperaquine for treating uncomplicated Plasmodium falciparum malaria. *Cochrane Database Syst Rev.* 2014(1), CD010927